

# NANOPORE SEQUENCING TO RESOLVE KIDD BLOOD GROUP DISCREPANCIES

M Gueuning<sup>1</sup>, L Schneider<sup>2</sup>, GA Thun<sup>1</sup>, N Trost<sup>2</sup>, S Sigurdardottir<sup>2</sup>, C Engström<sup>3</sup>, G Rizzi<sup>3</sup>, Y Merki<sup>2</sup>, K Neuenschwander<sup>2</sup>, C Gassner<sup>4</sup>, BM Frey<sup>1, 2, 3</sup>, MP Mattle-Greminger<sup>1\*</sup>, S Meyer<sup>2\*</sup>

<sup>1</sup>Department of Research and Development, <sup>2</sup>Department of Molecular Diagnostics and Cytometry, <sup>3</sup>Department of Immunohematology, Blood Transfusion Service Zurich, Schlieren, Switzerland

<sup>4</sup>Institute for Translational Medicine, Private University in the Principality of Liechtenstein, Triesen, Liechtenstein

\* equal contribution

www.blutspendezurich.ch



## BACKGROUND

- Constant increase in read accuracy of long-read sequencing → clinical diagnostics
- Nanopore sequencing has great potential for resolving complex phenotype/genotype discrepancies observed in routine blood group antigen typing
- Classical Sanger sequencing not adequate for:
  - detecting structural variants (SVs)
  - phasing novel variants to haplotype background
- Case example: Kidd system (Jk, *SLC14A1*)

## METHODS

- Routine donor genotyping by MALDI-TOF MS at Blutspende Zurich includes: *JK\*A/B* (c.838G>A), *JK\*01N.03* (c.582C>G), *JK\*02N.01/.02* (c.342-1G>A/C)
- Resolving 10 phenotype/genotype discrepancies (Fig. 1)
  - *SLC14A1* amplified in two overlapping long-range PCRs (Fig. 2)
  - Nanopore sequencing on a MinION (R9.4.1) flow cell
  - Sanger sequencing of exons and 'bridge-PCR' of breakpoints for validation

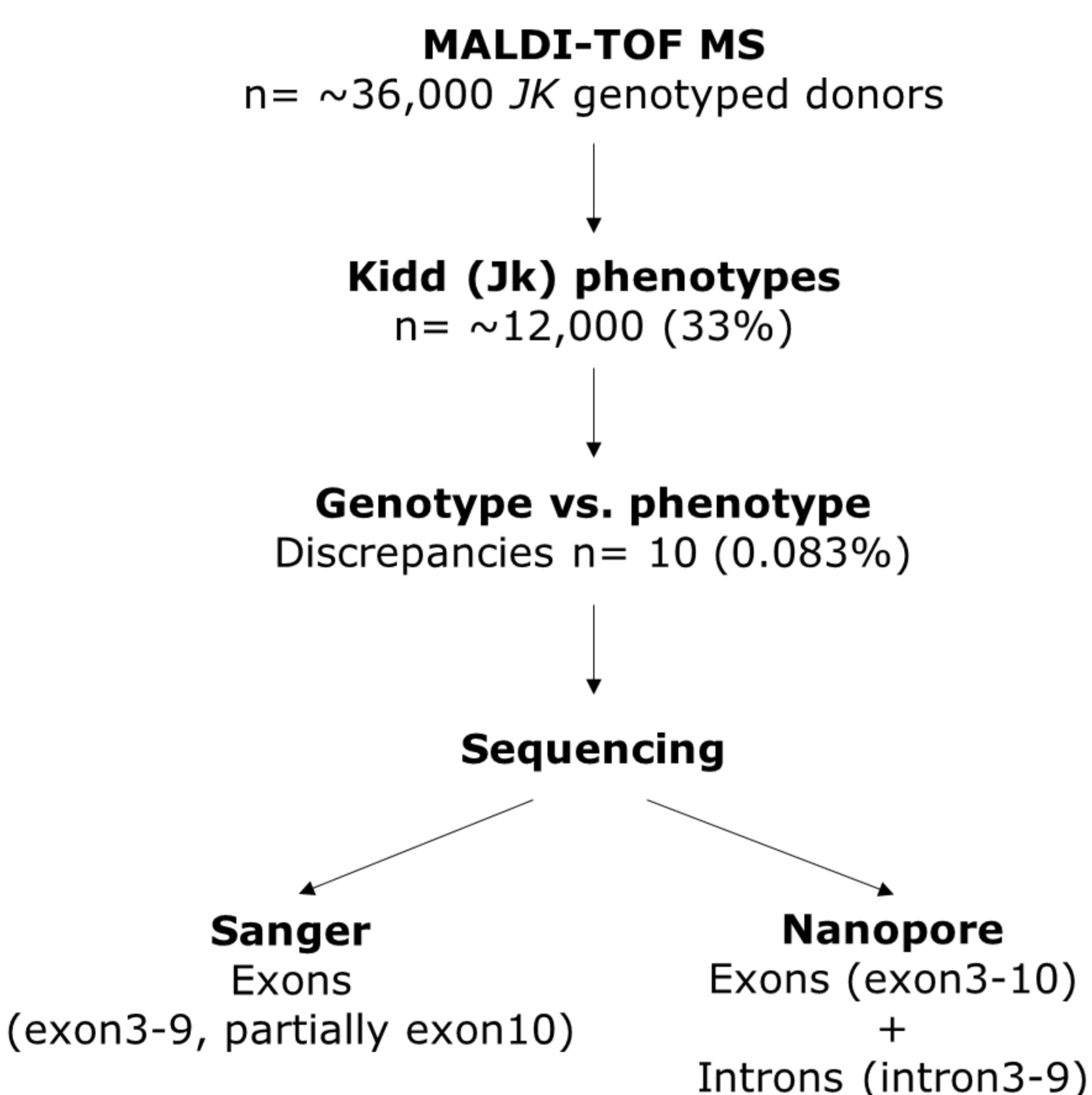


Figure 1. Flowchart of sample analysis.

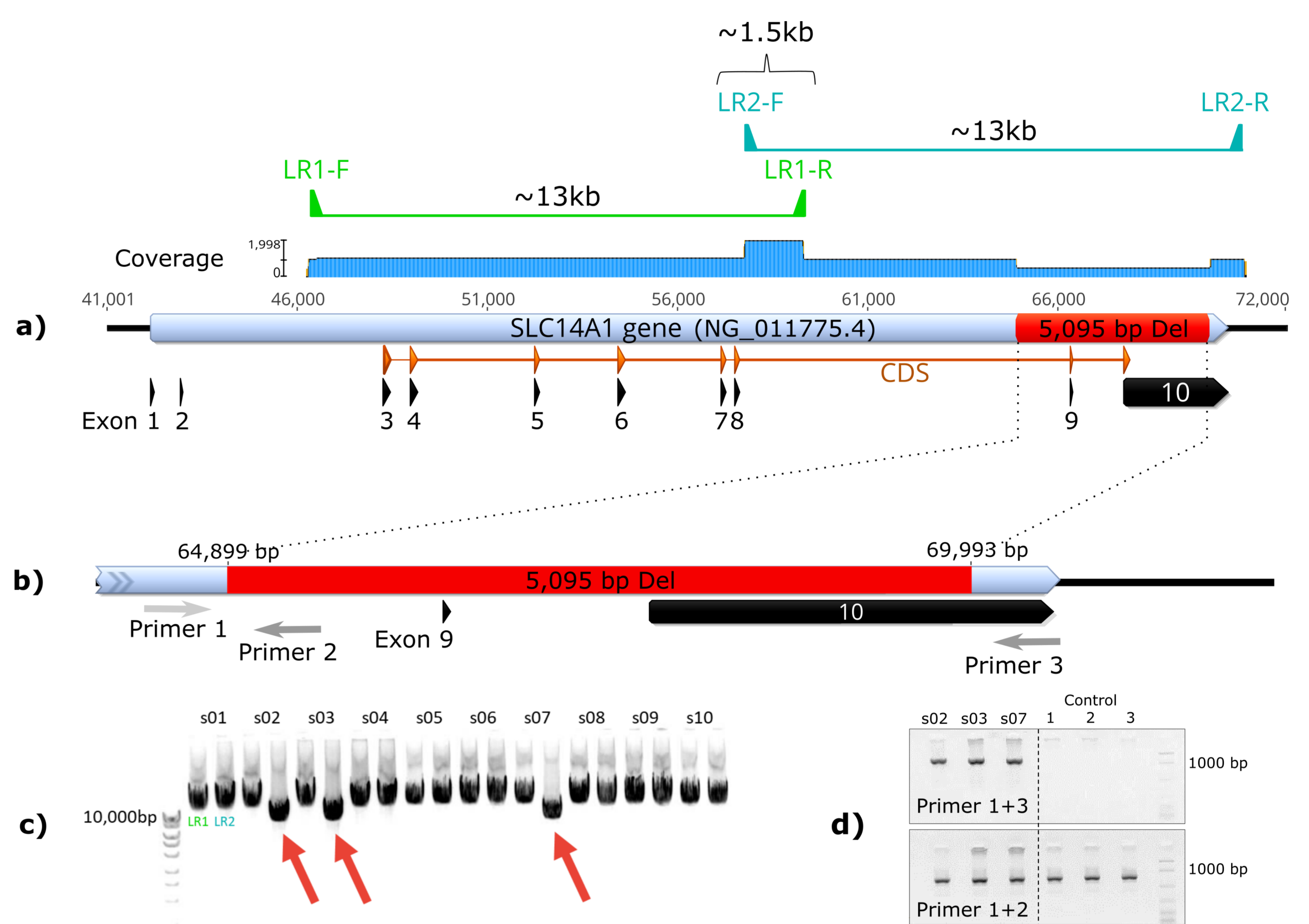


Figure 2. (a) *SLC14A1* gene showing exons, coding DNA sequence (CDS), newly identified ~5 kb deletion (red), coordinates of long-range PCRs (LR1/LR2), and raw reads mapping coverage of sample s02. (b) Zoom on the novel ~5 kb deletion with breakpoints and primer positions used for 'bridge-PCR'. (c) Long-range PCR products of all samples; red arrows highlight ~5 kb shorter LR2 fragments with novel deletion. (d) 'bridge-PCR' products for the three samples with the novel deletion, and control reactions.

## RESULTS & CONCLUSIONS

- 10 unexplained Kidd phenotype/genotype discrepancies among ~12,000 donors (Fig. 1)
- All discrepancies resolved using nanopore sequencing (Fig. 2, Table 1):
  - 5/10: rare known weak and null alleles not included in MALDI-TOF MS assay
  - 5/10: novel null alleles
    - SNV-based (n = 2)
    - *JK\*A* linked ~5 kb deletion (n = 3)
- Nanopore sequencing particularly useful for challenging diagnostics involving hybrid genes, deletions, and duplications
- Structural variants mostly not detectable by Sanger sequencing → likely more SVs missed so far also in other blood groups
- Established a fast and reliable protocol well adaptable to other blood group systems

Table 1. Summary table of serology, genotyping, and sequencing results for all 10 samples.

Sample	Serology	MALDI-TOF Genotyping	Nanopore Sequencing	
			Haplotype 1	Haplotype 2
<b>Known weak and null alleles</b>				
s01	Jk(a <sup>weak</sup> b-)	Jk(a+b+)	<i>JK*01W.01</i>	<i>JK*02N.08</i>
s04	Jk(a+b-)	Jk(a+b+)	<i>JK*01</i>	<i>JK*02W.04</i>
s05	Jk(a-b+)	Jk(a+b+)	<i>JK*01W.05</i>	<i>JK*02</i>
s06	Jk(a+b-)	Jk(a+b+)	<i>JK*01</i>	<i>JK*02N.09</i>
s08	Jk(a <sup>weak</sup> b-)	Jk(a+b+)	<i>JK*01W.01</i>	<i>JK*02N.06</i>
<b>Novel null alleles</b>				
s09	Jk(a+b-)	Jk(a+b+)	<i>JK*01</i>	<i>JK*02(G242E)Null</i>
s10	Jk(a+b+)	Jk(a+b+)	<i>JK*01(G40D)Null</i>	<i>JK*02</i>
<b>Novel structural variant</b>				
s03	Jk(a-b+)	Jk(a+b+)	<i>JK*01(Ex9_10del)*</i>	<i>JK*02</i>
s04	Jk(a-b+)	Jk(a+b+)	<i>JK*01(Ex9_10del)*</i>	<i>JK*02</i>
s07	Jk(a-b+)	Jk(a+b+)	<i>JK*01(Ex9_10del)*</i>	<i>JK*02</i>

\*Only detected by Nanopore sequencing. Confirmed by bridge-PCR (Fig. 2d).